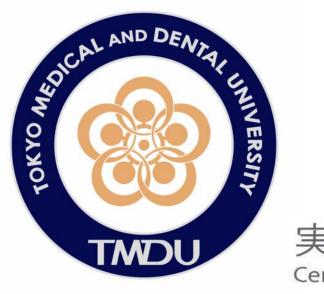
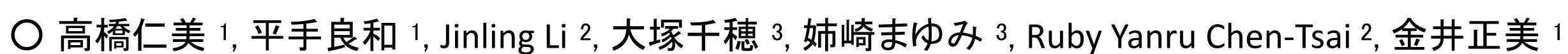
TARGATT™ノックインマウス技術を用いたサイト特異的トランスジェニックマウスの作製





Production of site-specific transgenic mice using the TARGATTTM knock-in technology







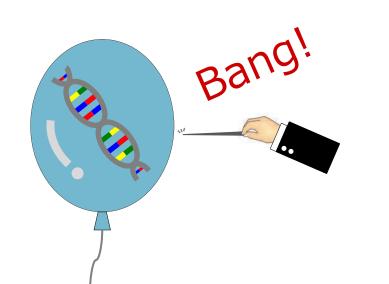
Summary

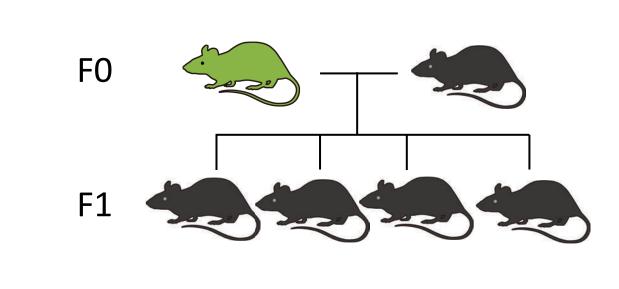
To produce transgenic (TG) mice, the technique of pronuclear injection of DNA, which results in random integration of the transgene, is commonly used. However, this classical method is unable to regulate the insertion site and copy number of the transgene, which occasionally causes disruption of endogenous genes by transgene insertion and silencing by tandem insertion of transgene. To overcome these problems, an US company, Applied Stem Cell Inc. (ASC), has developed the TARGATT™ TG system, which is based on φC31 integrase-based knock-in technique and has a great advantage in highly efficient, single-copy insertion in an active locus. ASC is developing TG mice and rats under the exclusive license from Stanford University. The φC31 integrase catalyzes an irreversible recombination event between triple tandem attP sites (attPx3) that have been engineered into a preselected, safe harbor locus (Hipp11) in the mouse genome, and a donor vector containing the gene of interest flanked by attP-recognition (attB) sequences. Co-injection of the donor DNA and φC31 integrase RNA into pronuclear stage embryos enables large transgene insertion (up to 22 kb) with very high efficiency (up to 40% or more). Our center has contracted a collaborative research agreement with ASC and are making TG mice using the TARGATT™ KI technology in consultation with ASC. We have already succeeded in the production of CAG-eGFP TG and Cre-ERT2 TG mice

Background & Purpose

Problems in common TG production

Disruption of an endogenous gene by the inserted transgene

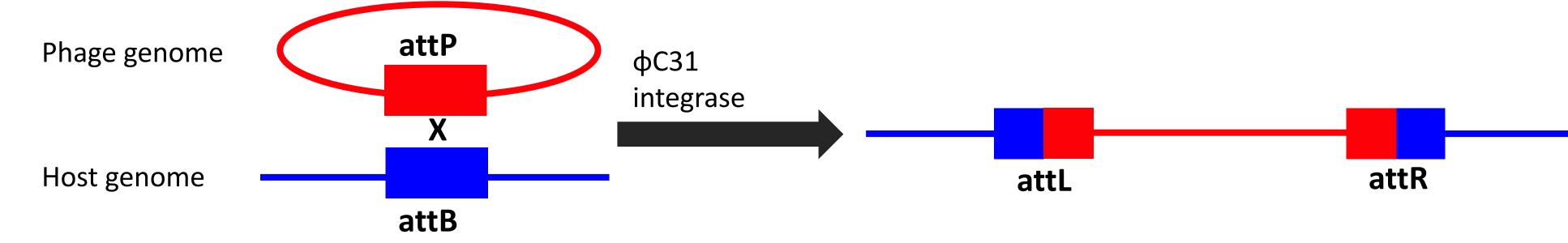




Gene silencing due to tandem insertion

φC31 integrase system is,

a site-specific gene integration system originally identified as a mechanism of phage DNA insertion into bacteria genome. The system requires no cofactors, therefore, it is active in other organisms, allowing us to use it as a tool for virus-free, one step integration of a gene of interest.



φC31 integrase,

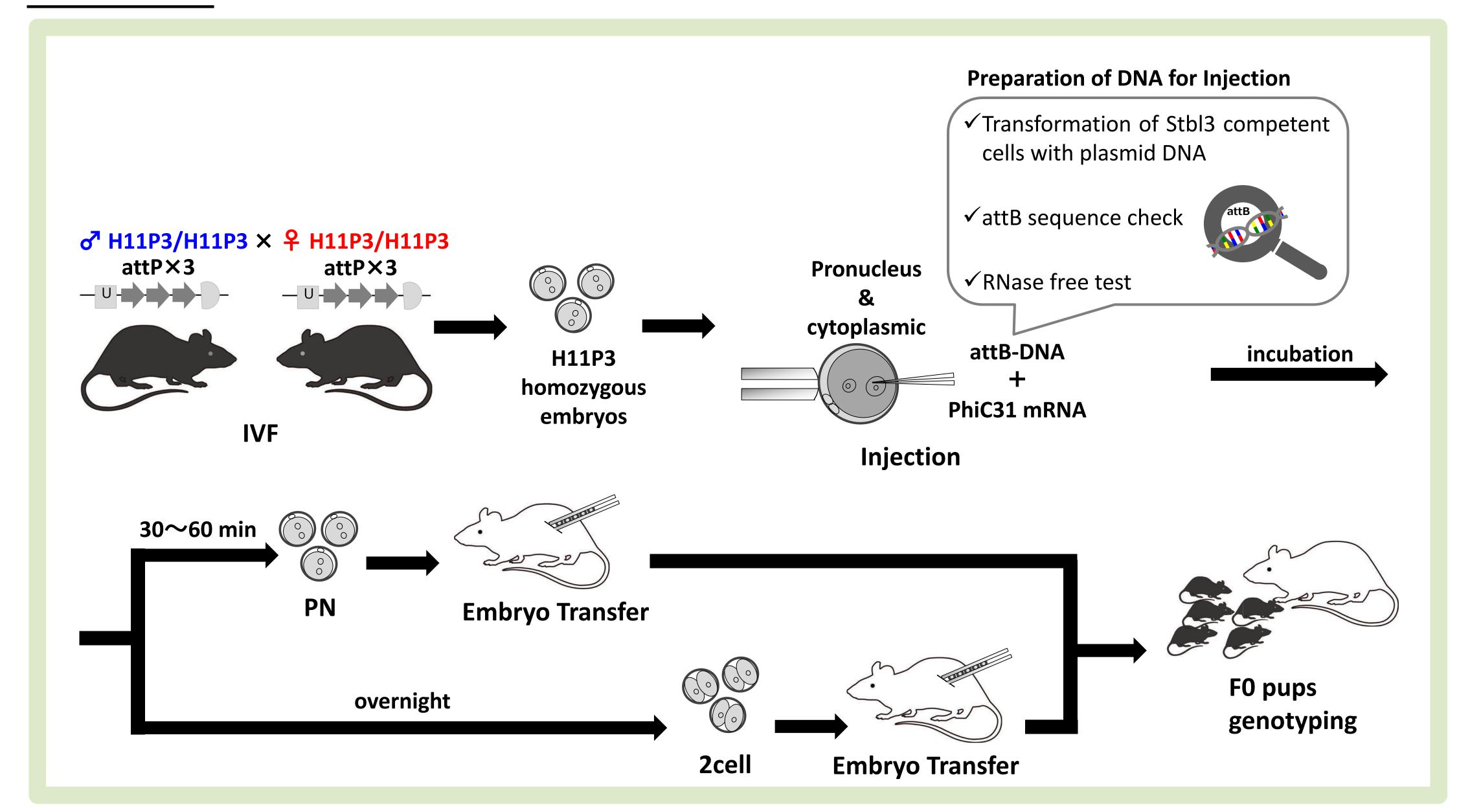
is a sequence-specific serine recombinase encoded in bacteriophage φC31 genome. It catalyzes recombination between attP sequence in the phage genome and attB sequence in the host genome, which results in integration of attL-attR in the host genome.

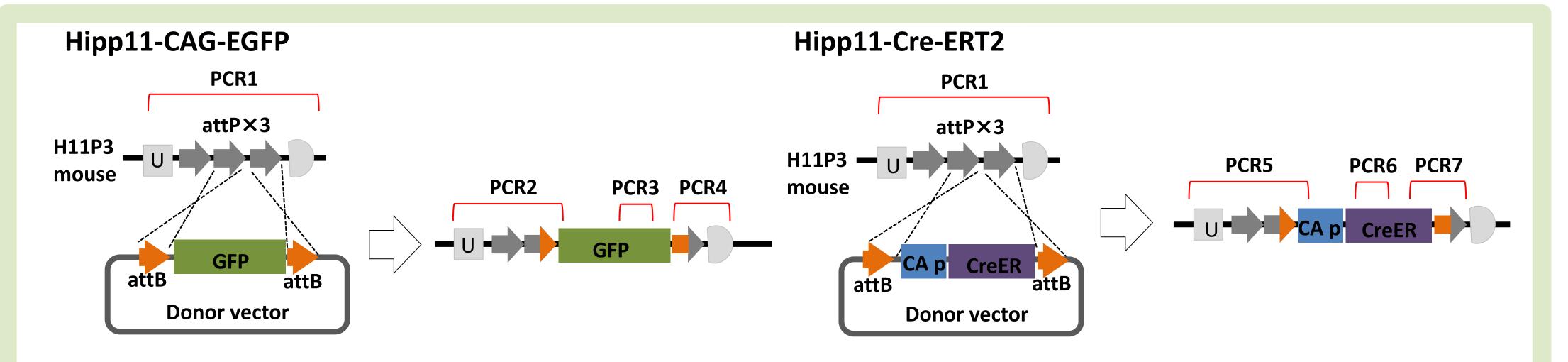
> TG mice production using φC31 integrase system

(TARGATT™ knock-in system)

The φC31 integrase catalyzes an irreversible recombination event between triple tandem attP sites (attPx3) that have been engineered into a preselected, safe harbor locus (Hipp11) in the mouse genome, and a donor vector containing the gene of interest flanked by attP-recognition (attB) sequences. Co-injection of the donor DNA and φC31 integrase RNA into pronuclear stage embryos enables large transgene insertion (up to 22 kb) with very high efficiency (up to 40% or more). Hipp11 is a locus in chromosome 11, which stably induce the integrated gene like ROSA26 locus.

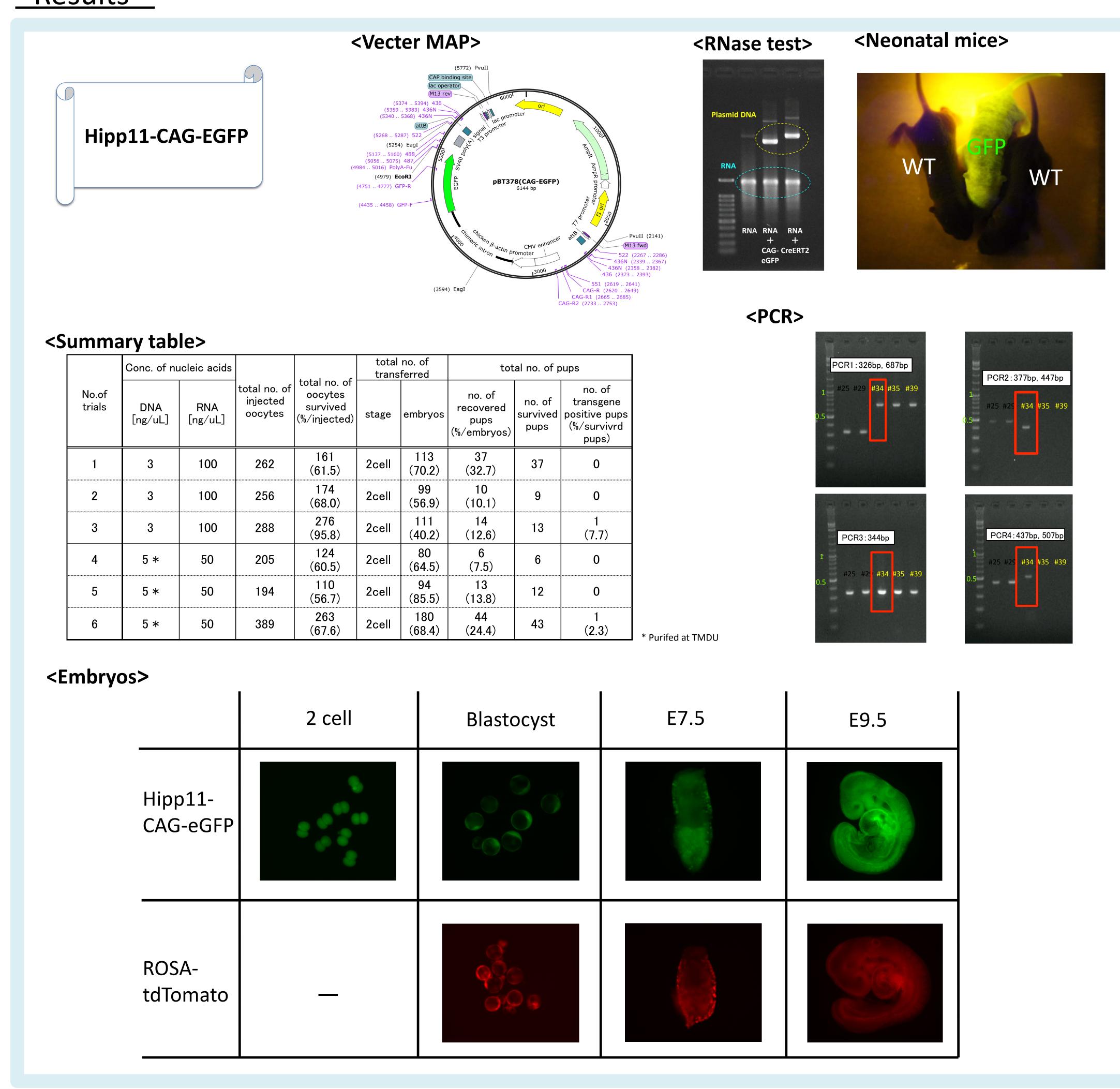
Method

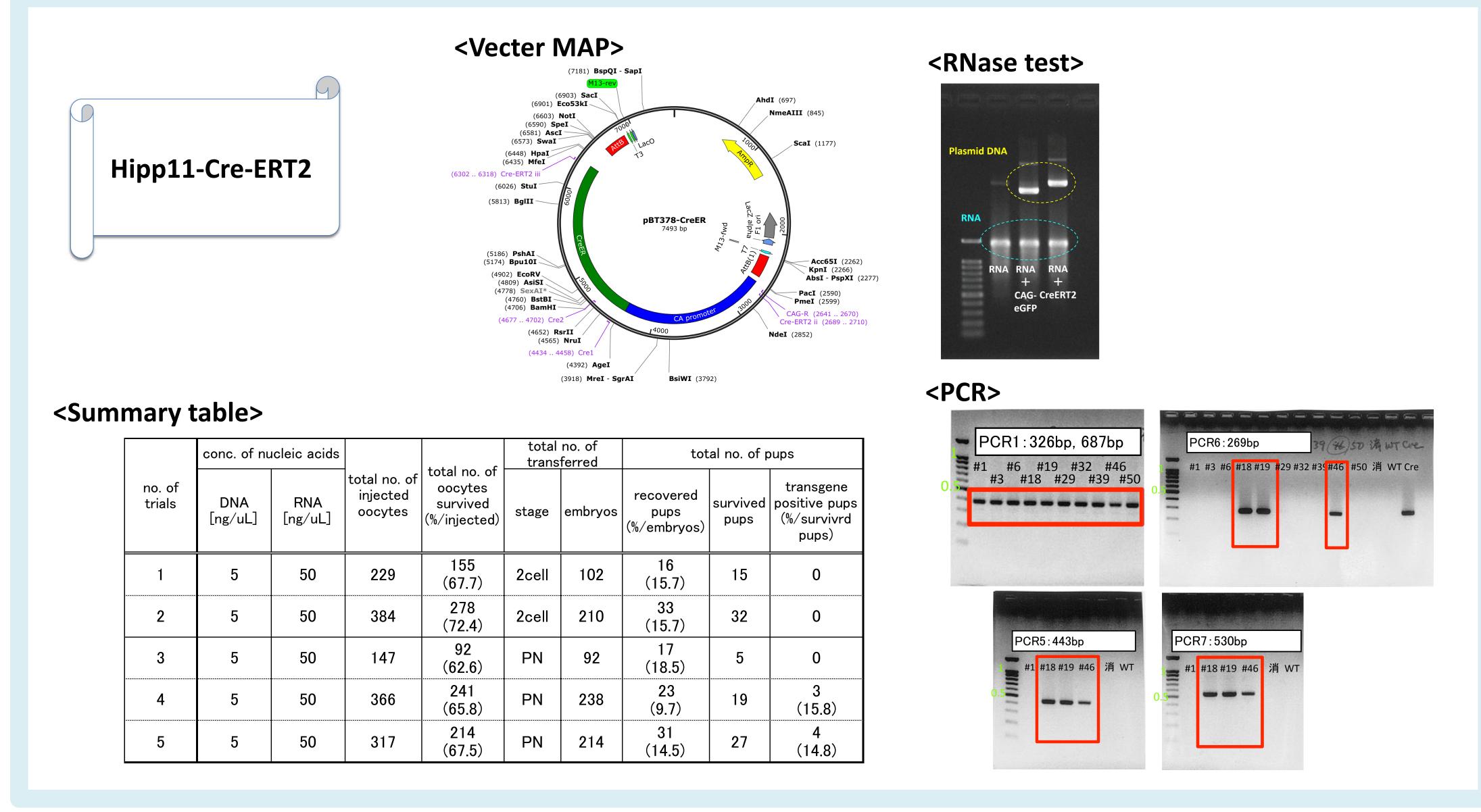




Results

O Hitomi Takahashi ¹, Yoshikazu Hirate ¹, Jinling Li ², Chiho Otsuka ³, Mayumi Anezaki ³, Ruby Yanru Chen-Tsai ², Masami Kanai-Azuma ¹





Discussion

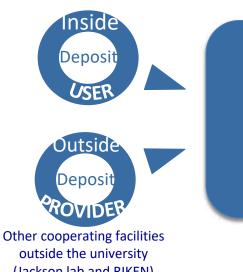
1. Advantages in TARGATT™ TG mice production

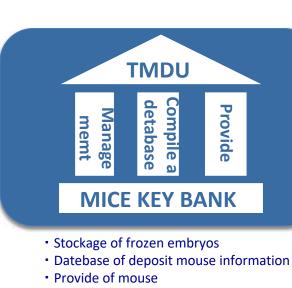
- Do not disrupt endogenous gene expression
- Time-saving and cost-effective gene integration using highly efficient φC31 integrase system
- Single copy knock in is free from repeat-induced gene silencing and genomic instability
- No need of large-scale clone screening
- Variety of donor plasmid vectors TARGATTTM2: CAG-PolyA, TARGATTTM3: no promoter-MCS, TARGATTTM6.1: CAG-L4SL-MCS-PolyA, TARGATTTM7: PGK-MCS-PolyA,

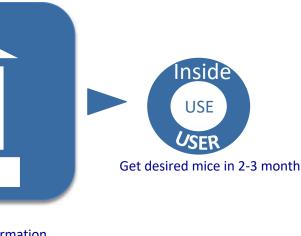
2. The Mice Keybank system at TMDU: Storage and resource system of TARGATT ™ strains for internal

Our Center is planning to provide the produced TARGATT TGs to the institutional internal users via the Mice Keybank system, which was founded to collect, store and share useful mouse strains among internal users.

TARGATTTM8: PCA-MCS-PolyA, TARGATTTM9.1: PCA-L4SL-MCS-polyA







Acknowledgment

We thank Prof. Hiroshi Asahara and Dr. Yoshiaki Ito (Dept. of Systems BioMedicine, TMDU) for construction of Cre-ERT2 plasmid.